

FreeAdd 1X

CAT N°: SFA-500

Theoretical Ph: 7.5 ± 0.5

Colour: Light opalescent liquid

Storage conditions: +2°C to +8°C, protected from light

Shelf life: 12 months

Sterility tests:

- bacteria in aerobic and anaerobic conditions
- fungi and yeasts

Endotoxin: < 7.5 EU/ml

Recommended use:

- Respect storage conditions of the product
- Do not use the product after its expiry date
- Store product in an area protected from light (not necessary for saline solutions).
- Manipulate the product in aseptic conditions (e.g. : under laminar air flow)
- Wear clothes adapted to the manipulation of the product to avoid contamination (e.g. : gloves, mask, hygiene cap, overall...)

The product is intended to be in vitro laboratory used or for further manufacturing in vitro used only.
Do not use it in therapy, human or veterinary applications.

Applications:

FreeAdd is a **chemically defined substitute for animal serum**.

It provides the necessary nutritional support for cell growth, development and expression.

FreeAdd stands for a cell culture supplement designed to enable the in vitro culture of animal cells in a chemically defined, animal-derived component free environment.

FreeAdd replaces most supplements like Fetal Bovine Serum in an attempt to reproduce the normal extracellular environment, the extracellular fluid, making your culture experience predictable, safe and future-proof. It lowers costs for downstream processing due to near absence of proteins. Being fully defined there is no need for batch testing and batch storing, and it can be stored cool.

The use of FreeAdd is appropriate for most animal cell types, both anchorage-dependent and suspension cell culture types, and supports the growth and maintenance of cell lines as well as primary cultures.

Cells grown in the absence of animal sera are in general more sensitive to antibiotics, enzymes, hormones and growth factors. Hence, these components should be adjusted accordingly. A 1.5 – 2-fold decrease in concentration represents a good starting point.

The use of non-tryptic cell dissociation and non-mammalian reagents such as Accutase (Cat No SLA-590-100) is recommended.

General instructions for use :

Add FreeAdd 1X to your basal cell culture medium of choice. For the great majority of cell types a 10% concentration is indicated. The complete medium is stable for 30 days when stored in the dark at +2/+8°C.

Before use, let FreeAdd equilibrate to room temperature or heat to 37°C in a water-bath with occasional gentle swirling.

Infrequently some flocculent material may appear, but this material will go into solution with gentle swirling at 37°C.

Cell lines adapted to growth in the presence of animal sera such as FBS can be converted to serum-free growth by following basically two protocols:

- a) direct adaptation by shifting the serum-containing cell culture medium to FreeAdd supplemented culture medium in one single step or
- b) through a less harsher procedure by progressively phasing out the serum content

IMPORTANT NOTES BEFORE USING FreeAdd:

- FreeAdd is a cell culture medium additive replacing serum, so it is not a final medium.
- FreeAdd must be added to a basal cell culture medium (e.g. IMDM, DMEM-F12 or any other basal medium of choice).
- Do not filter FreeAdd or the medium after adding FreeAdd.
- If needed filter your medium first and then add FreeAdd under sterile conditions.
- FreeAdd does not contain growth factors like cytokines, hormones etc. Therefore also no insulin.

Ideally, the transition to serum-free conditions should be carried out over several passages to gradually select cells that can grow under serum-free conditions. However, direct adaptation to serum-free environments may also work out successfully, provided that all crucial aspects are addressed properly. Regardless of the method used, key concerns include the growth state of the cellular inoculum, cell seeding density, sub-cultivation techniques, and biophysical attributes of the cell culture system. FreeAdd serum replacement has been designed so as to be used in the same way as conventional cell culture sera, as a medium supplement.

Detailed instructions for use:

1 Preparing

1.1 General preparation of the serum-free cell culture medium

Gently shake the bottle of FreeAdd shortly before use.

Add FreeAdd 1X to your preferred basal medium at the same concentration compared to serum (e.g. 10%).

Do not filter FreeAdd or the medium after adding FreeAdd (FreeAdd is sterile).

If needed filter the medium before adding FreeAdd.

Do not add any antibiotics at this stage. In fact, antibiotics like many compounds bind to the plasma proteins of serum, in particular to the albumin fraction. Thus, the same concentration of antibiotics will exhibit a much higher biological activity in serum- and albumin-free conditions and this increased activity may have deleterious impacts on cell growth.

In case 'antibiotic-free culture' is deemed unworkable, the use of gentamycin is suggested at the concentration of 50 mg/l.

**Some users prefer or need to grow their cells free of insulin. In case of FreeAdd the decision to grow cells with or without insulin (and/or other growth factors) is yours. If insulin is needed for cell growth and performance we advise to add recombinant insulin in a concentration of 1.25 mg/l final cell culture medium.*

1.2 General adaption methods to serum-free conditions

There are basically two approaches to adapt cells to growth in serum-free environment:

1.2.1 Direct Adaptation

Which is carried out by a direct transfer of the cells from the serum-containing medium into the serum-free medium.

1.2.2 Sequential Adaptation or Weaning Method

Pass the cells from the original serum containing medium sequentially through the following phases where each step halves the serum-supplemented media, thus increasing the serum-free media to approximately below values:

Phase 1: 50 % FreeAdd-supplemented medium	/	50 % Serum-supplemented medium
Phase 2: 75 % FreeAdd-supplemented medium	/	25 % Serum-supplemented medium
Phase 3: 87.5 % FreeAdd-supplemented medium	/	12.5 % Serum-supplemented medium
Phase 4: 93.75 % FreeAdd-supplemented medium	/	6.25 % Serum-supplemented medium
Phase 5: 96.88 % FreeAdd-supplemented medium	/	3.12 % Serum-supplemented medium
Phase 6: 98.44 % FreeAdd-supplemented medium	/	1.56 % Serum-supplemented medium
Phase 7: 100 % FreeAdd-supplemented medium		

In case of reduced growth go back one step and continue after growth is established again.

Cell cultures may consist of cell lines (adherent or suspension growth) or primary cultures. Moreover, from a functional point of view, cell types may be differentiated to various degrees or exhibit undifferentiated characteristics, as in the case of stem cell preparations.

In each case, the adaptation protocol has to take into account the specific requirements of the cell type in order to guarantee the best chances for success.

2 Specific adaptation procedures

2.1 Cell lines

The following protocols are valid for normal (diploid, limited lifespan) or transformed or immortalized cell lines (with indefinite lifecycle).

2.1.1 Anchorage-dependent Cell Lines

Critical success factors:

- coating of the cell culture support for optimal cell attachment
- minimize action of trypsin
- choice of the antibiotic system

Experimental Steps:

A) Coat the cell culture surface with an adequate cell-attachment factor by using

- a commercial coating kit such as Pronectin™ F, MapTRIX™ or equivalent, or
- a Fibronectin or Poly-L-Lysine coating, or
- ready to use plastics that provide an improved attachment of adherent cells

B) Dissociate the cell monolayer

Biowest advises to employ a non-mammalian dissociation reagent such as Accutase (Cat No SLA-590-100) which does not require inactivation or removal during passaging. Accutase (Cat No SLA-590-100) dissociates the cell monolayer; this cell detachment solution has been developed to meet the most demanding requirements for gentle and effective detachment of adherent cells; cell membranes and surface epitopes will not be harmed and the structural and functional quality of the surface proteins remain intact.

C) Seed cells at 20,000 cells per cm² in complete medium as prepared under point 1.

It is important to observe a high seeding density during the first steps of the adaptation process. Cells normally secrete a host of factors into the culture medium that control cell attachment, growth and proliferation. However, during the seeding step these factors are absent in the fresh serum-free medium and a critical level of cell density is essential to induce an immediate and sufficient production of these autocrine/paracrine factors.

D) Incubate and maintain the cell cultures at 37° C until they reach 80-90% confluency.

During this period change 75% of the medium every 2 -3 days. Do not discard the spent medium. Instead harvest the conditioned medium, sterile filter and put aside at 4°C for use in the next steps. If the cells seem stalled at any point, allow them more time to adapt to their new serum-free environment.

E)) When near confluency is reached, split the cells at a 1:2 or 1:3 ratio.

For this second passage in FreeAdd a coating is not required but use of conditioned medium is strongly suggested – this medium fraction contains indeed the autocrine factors that regulate attachment, spreading, growth and proliferation.

Seed cells in a mixture consisting of 75% fresh medium + 25% conditioned medium, collected during the previous passage.

Continue supplying cells with 75% fresh medium every 2-3 days and collect the conditioned medium as under d) above.

F) Repeat step E) until the cells exhibit growth dynamics comparable to their former growth in serum-supplemented medium. At that point the cell line can be considered fully adapted. This may take up to a total of 4-6 passages.

G) From this point on, antibiotics may be added to the culture medium. We advise the use of the large-spectrum antibiotic gentamycin; this antibiotic has a much reduced cytotoxicity as compared to the

standard Penicillin/Streptomycin cocktails. The suggested concentration of use of gentamycin is 50 mg/l (Cat No SLG-504-100).

H) Once adapted, the original split ratio (in serum-supplemented conditions) may be applied.

2.1.2 Anchorage-independent Cell Lines

The following protocol is adjusted for cell lines that grow already in suspension.

Critical success factor:

Choice of the antibiotic system

Experimental Steps

A) When cell densities of $3-5 \times 10^6$ cells/ml are reached (depending on the cell line) start switching to FreeAdd supplemented medium. Harvest the cell suspension, take out a small aliquot for cell counting and centrifuge the whole suspension at 200 g for 5 minutes.

B) Perform a cell count.

C) Resuspend the cell pellet in FreeAdd supplemented medium at a density of 10^6 cells/ml.

It is important to observe a high seeding density during the first steps of the adaptation process. Cells normally secrete a host of factors into the culture medium that control cell growth and proliferation. However, during the seeding step these factors are absent in the fresh serum-free medium and a critical level of cell density is essential to induce an immediate and sufficient production of these autocrine/paracrine factors.

D) Incubate and maintain the cell cultures at 37°C until they reach a density of approximately $3-5 \times 10^6$ cells/ml.

E) Split the suspension cultures at a 1:3 or 1:4 ratio, by adding the appropriate volume of fresh medium (e.g. 25 ml of cell suspension + 75 ml FreeAdd supplemented medium, to be dispatched into 4 separate culture vessels)

F) Repeat step E) until the culture exhibits growth dynamics as originally in serum-supplemented medium.

From then on, the cell line can be considered fully adapted and may be split at the original ratios during serum-supplemented culture.

G) From this point on, antibiotics may be added to the culture medium.

We suggest the use of Gentamycin at the concentration of 50 mg/l (Cat No SLG-504-100); this antibiotic has a much lower cytotoxicity as compared to the standard Penicillin/Streptomycin cocktails.

2.2 Stem Cells

2.2.1 Preliminary step - Coating of the culture surface

When growing human pluripotent stem cells (hPSCs) or multipotent mesenchymal/stromal cells (MSCs) in defined, feeder-free systems the treatment of the culture surface with an adequate coating strategy is of crucial importance. Usually crude preparations of extracellular matrices, such as Matrigel™ (BD Biosciences) are commonly used.

However, the undefined nature (mouse tumor-derived) as well as the presence of animal-derived compounds renders the use of Matrigel problematic in applications where the clinical potential of hPSC is pursued.

In that case the use of another coating agent, StemAdhere™ is recommended. StemAdhere™ is a defined matrix containing a single recombinant protein composed of entirely human sequences and can therefore be defined as Animal Component-Free (ACF). However, one disadvantage of

StemAdhere™ comes from the fact that non tissue culture-treated plates are to be used for the coating with StemAdhere™ Defined Matrix.

A third feeder free, xeno free and chemically defined alternative is represented by Synthemax™ Surface from Corning. This product has been designed to mimic a cells' natural environment and has the advantage to be offered as specially treated, ready-to-use culture plasticware. It has a proven track record of good results for several hESC and hiPSC lines. When switching to Synthemax™ an initial adaptation period may be noted, but after several passages the cells look perfect again.

It is not within the scope of the present 'Instructions for Use' to replicate the details of the coating protocols for Matrigel™ and StemAdhere™.

Detailed information regarding Synthemax™ culture surface plastics can be found here:

http://www.corning.com/lifesciences/us_canada/en/technical_resources/surfaces/cell_culture/synthemax.aspx

IMPORTANT NOTES

- FreeAdd does not contain any growth factors and therefore no bFGF and insulin. We recommend to add bFGF and insulin or IGF when culturing stem cells
- FreeAdd does not contain selenium. We advise to use a selenium containing medium when culturing stem cells

2.2.2 Feeder-Free Culture of hESCs and hiPSCs

Pluripotent stem cell colonies may be cultured feeder-free in 2-3% FreeAdd-supplemented DMEM/F-12 medium with a prior coating step as outlined above.

It has been established that extrinsic and autocrine signaling are responsible for matrix remodeling and maintenance of embryonic stem cell renewal (Przybyla, L.M. and Voldman J. PNAS vol. 109 no. 3, 835-840, 2012). For this reason, and in order to not deplete the cultures of these important factors, it is of highest importance to strictly adhere to the medium change protocol as described below.

Preparation of the complete culture medium for hESCs and hiPSCs

- Use your conventional basal culture medium (e.g. DMEM-F12)
- Add L-Glutamine to a concentration of 2 mM (e.g. 1.0 ml of a 200 mM stock solution Cat No X0550 for a final volume of 100 ml of medium)
- Add bFGF to a final concentration of 4 ng/ml (e.g. 40 µl of a stock solution of 10 µg/ml for a final volume of 100 ml of medium)
- Add 2-mercaptoethanol to a final concentration of 0.1 mM (e.g. 182 µl of a stock solution of 55 mM for a final volume of 100 ml of medium)
- When opting for an antibiotic protection system, we suggest to use gentamycin at the concentration of 50 mg/l (Cat No L0012)
- Since FreeAdd does not contain insulin, you may decide to add (recombinant) insulin or IGF to the medium
- Sterile filter the medium if needed
- Add 10% of FreeAdd 1X

Culture of hESCs in FreeAdd-supplemented medium – first passage

- Coat 6-well tissue culture plates with either Matrigel™ or StemAdhere™ or use Synthemax™ culture surface plastics
- Thaw a fresh ampoule of hESC or remove hESC cells from an existing culture (from a feeder or feeder-free culture, as adequate) using collagenase treatment or better, using Accutase and sediment as usual. Accutase is of non-mammalian origin and has both protease and collagenolytic activity; it has shown outstanding performance with hESC cells
- Matrigel™ or StemAdhere™ coated plates: Aspirate the excess Matrigel or StemAdhere coating agent
- Synthemax™ culture plates: no action needed, use as such
- Plate the triturated colonies in complete medium prepared as described above

- Feed cells every day up to 7 days by changing 75% of the medium; it is of key importance to leave 25% of the medium containing the autocrine factors produced by the cells

Note: Colonies can grow bigger and more densely on Matrigel™ without losing morphology than on MEFs.

Passaging of the cell cultures

- Wash cells once with Dulbecco's Phosphate Buffered Solution (DPBS).
- Add dispase (e.g. 1 ml/well of a 2 mg/ml enzyme solution in e.g. DMEM-F12) and incubate at 37°C.
- Colonies should detach intact within 10–15 minutes upon tapping smartly on the side of the plate.

Attention: DO NOT SCRAPE

- Transfer the dispase solution containing the colonies into a sterile 15 ml tube and rinse the culture wells with an additional 1ml/well of growth medium in order to harvest all colonies.
- Centrifuge and wash twice more as usual.
- Triturate very gently by pipetting as colonies grow flat and dissociate readily in dispase.

Note: Small colonies and single cells do not survive well.

- Plate at the usual split ratio on coated plates (Matrigel, StemAdhere, Synthemax - please see above).

2.2.3 Culture of MSCs

The following protocol is intended for culturing MSCs in defined conditions, starting from a frozen ampoule from a liquid nitrogen stock or from a culture growing in a different cell culture system.

General considerations

- MSCs should be stored in liquid nitrogen if not seeded directly after derivation. Storage at higher temperatures (-80°C) may cause irreversible cell damage.
- Use aseptic techniques and work in a laminar flow hood.
- Incubate cells in a humidified incubator at 37°C, 5% CO₂.
- The cultureware must be treated by a coating step as outlined in the preliminary chapter of part 2.2.1 of these Instructions for Use "Coating of the culture surface".
- Always seed cells at a density of 2000 cells per cm². Avoid growing the cells to confluency. Subculture the cells when a density of approximately 70% is reached.
- Use dissociating enzymes that do not need to be inactivated with serum, such as Accutase (Cat No SLA-590-100).
- After harvesting, always re-suspend the cell pellets by gentle pipetting. Never vortex the cells.
- Prepare all required materials and equipment before performing any of the culture processes described below.
- Always pre-warm all solutions and media which come into contact with the cells.

Preparation of the complete culture medium

- You can use your conventional basal culture medium.
- Add L-glutamine to a concentration of 2 mM (e.g. 1.0 ml of a 200 mM stock solution Cat No SRL-811-100 for a final volume of 100 ml of medium).
- Add bFGF to a final concentration of 4 ng/ml (e.g. 40 µl of a stock solution of 10 pg/ml for a final volume of 100 ml of medium).
- When opting for antibiotic protection system, we suggest to use gentamycin at the concentration of 50 mg/l.
- Since FreeAdd does not contain insulin, you may decide to add (recombinant) insulin or IGF to the medium.
- Sterile filter the medium if needed.
- Add 10% of FreeAdd 1X.

Thawing of cells

During the thawing stage, care must be taken to handle cells gently and placing them immediately into pre-warmed complete culture medium.

- Prepare a 15 ml conical centrifugation tube containing 10 ml pre-warmed complete medium.
- Remove cells from liquid nitrogen storage.
- Place vial of cells in 37°C water bath and agitate moderately until all ice has melted.
- Immediately disinfect the vial with 70% ethanol.
- Transfer cells immediately into the centrifugation tube containing the pre-warmed medium and spin at 300 g for 5 min.
- Aspirate the supernatant and carefully re-suspend the cell pellet in complete medium.
- Seed the cells at a density of 2000 to 4000 cells per cm² in a cell culture dish coated with either Matrigel™ or StemAdhere™ or in ready-to-use treated Synthemax™ culture dishes (Greiner).
- Incubate cells at 37°C in a 5% CO₂ humidified incubator.
- Feed the cells every day by changing 75% of the medium; it is of key importance to leave 25% of the medium containing the autocrine factors produced by the cells.

When approximately 70% of confluency is reached passage the cell cultures.

Subculturing the cells

- Aspirate the cell culture medium and wash the cells once with Dulbecco's Phosphate Buffered Saline w/o Calcium w/o Magnesium (Cat No SLD-576-500).
- Submerge cells with a sufficient volume of Accutase (Cat No SLA-590-100) solution and incubate for 5 minutes at 37°C.

If needed, detach cells by softly tapping the side of the cell culture vessel.

- Add complete cell culture medium to dilute the enzyme solution (add at least twice the volume of Accutase).
- Transfer the cell suspension to a centrifugation tube and spin down the cells for 5 min at 300 g.
- Discard the supernatant and resuspend the cells in complete medium by cautiously pipetting up and down.
- Perform a cell count.
- Seed the cell suspension in new coated cell culture dishes or Synthemax™ dishes (see "Coating of the culture surface" above) at a density of 2000 cells per cm².
- Incubate cells at 37°C in a 5% CO₂ humidified incubator.
- Proceed to the next subculture pass as soon as a cell density of approximately 70 % is reached. Subculturing is usually required twice a week.

2.3 Primary Cultures

Primary cell cultures consist in growing cells immediately after their isolation from a living tissue or organism. They represent the core of the cell culture world: all existing cell lines to date have been initiated as primary cultures and this paradigm is here to stay.

But apart from generating new cell lines, primary cultures represent also a very important tool by themselves, especially in fields such as drug discovery and development, regenerative medicine and fundamental research.

From a technical point of view, primary cell cultures remain also the most delicate part in the cell culture process. Freshly isolated cells have not been submitted to any selective pressure and remain highly representative of their in vivo counterpart. This fact calls for the outmost attention to satisfy their nutritional and physiological requirements. Ideally, the cell culture environment for primary cultures should mirror as closely as possible the in vivo situation, i.e. the extracellular space. This can only be achieved in defined cell culture conditions that allow for full control of the supply of nutrients and growth factors.

Primary cell cultures have highly disparate requirements, depending on their tissue of origin. FreeAdd has proven successful in growing various primary cell cultures in the absence of undefined additions such as bovine serum or its derivatives. There is however no universal recipe that would satisfy the needs of all primary cell types.

The following guidelines fall into two parts: general recommendations that apply to all cell types and specific requirements for the main tissue types.

2.3.1 Common recommendations

When opting for a serum-free primary cell culture process the following points need to be addressed, no matter which cell type is used.

ABSENCE OF SERUM ATTACHMENT FACTORS

Preliminary step - Coating of the culture surface:

In defined culture conditions, the treatment of the culture surface with an adequate coating strategy is of crucial importance.

Usually crude preparations of extracellular matrices (ECM), such as mouse sarcoma extracts (e.g. matrigel) or extracted collagen preparations are commonly used. However, the undefined nature as well as the presence of animal-derived compounds renders their use problematic for many applications.

In the case that a contact with animal-derived material does not pose a problem, a 'quick-fix method' consisting of an overnight treatment of the plastic cell culture surfaces with a small amount of FBS may be considered. This method is cost convenient and efficient, however it represents a back step from the fully defined culture environment concept.

Today recombinant and defined coating kits are available, that mimic the attachment properties of ECM proteins through the use of biosynthetic signaling peptides derived from fibronectin, laminin, collagen, E- cadherin, vitronectin etc.

ABSENCE OF ENZYME INHIBITORS FROM SERUM

Dissociation enzyme

There are essentially two ways to start a primary culture: by outgrowth from a primary explant or by enzymatic disaggregation.

In the latter method the starting tissue is digested by using proteolytic enzymes or cocktails of enzymes, such as dispase, collagenase and trypsin.

Care must be taken to neutralize / deactivate any remaining proteolytic activity before seeding the cells. This point must be addressed in particular when trypsin is employed.

The use of standard trypsin preparations can become problematic in the absence of serum, which contains trypsin inhibitors. In serum-deprived conditions the tryptic activity must be inactivated after the cell dissociation process, this can be achieved by using an efficient trypsin inhibitor, such as a soybean trypsin inhibitor.

As an alternative to trypsin, the use of Accutase is highly recommended because it does not need to be de-activated. This non-mammalian enzyme has been efficiently used for a series of primary cultures, including primary smooth muscle cells, primary human endothelial cells, primary chick neuronal cells.

2.3.2. Celltype-specific recommendations

Primary cell cultures have different cell culture requirements, depending on their tissue of origin.

In this booklet we will not detail the primary cell culture procedures that differ vastly from one cell type to the other. As a general rule we recommend to apply the 'conventional' techniques for the isolation of the primary cells of the desired type and REPLACE the serum contribution by adding 1x lower concentration of FreeAdd 1X.

This will satisfy the nutritional requirements of most, if not all cell types. Indeed, in the realm of mammalian cell cultures, nutritional requirements vary only slightly quality-wise, more demanding cell types, such as hepatocytes requiring higher nutrient concentrations.

The growth factor and hormone requirements however differ between cell types. The following table lists the cell culture media preparations that we recommend when FreeAdd is used as a replacement for animal serum.

The growth factor and hormonal additions are those indicated for optimal cellular development and proliferation with respect to each of the four indicated cell types.

<i>Primary Cell Culture Type</i>	<i>Recommended Growth Factor</i>	<i>Recommended Hormones</i>	<i>Final Medium Concentration</i>	<i>Requirement</i>
Primary kidney cultures				
FreeAdd 1X, 10%		Insulin (recombinant human)	0.5 µg/ml	essential
Basal medium : DMEM high glucose F-12 (ref.BW: SLD-515-500)		Hydrocortisone	0.1 µg/ml	essential
		Epinephrine	0.5 µg/ml	essential
	EGF (human, recombinant)		50 ng/ml	optimal/beneficial
		Triiodo-L-thyronine	10 pg/ml	essential
	EGF (human, recombinant)		10 ng/ml	optimal/beneficial
Primary hepatocytes				
FreeAdd 1X, 10 - 15%		Insulin (recombinant human)	5 µg/ml	essential
Basal medium : William's Medium E (available on request)		Hydrocortisone	0.5 µg/ml	essential
	EGF (human, recombinant)		50 ng/ml	optimal/beneficial
Primary keratinocytes*				
FreeAdd 1X, 10%		Bovine Pituitary Extract (BPE)	4 µl/ml	essential
DMEM / F-12 (ref.BW: SLD-515-500) 1:3 ratio		Hydrocortisone	5 µg/ml	essential
		Epinephrine	0.5 µg/ml	essential
	EGF (human, recombinant)		0.125 ng/ml	optimal/beneficial
Primary cardiomyocytes				
FreeAdd 1X, 10%		T3 (triiod-L-thyronine)	1 ng/ml (1.5nM)	essential
Claycomb Medium		Insulin (recombinant human)	5 µg/ml	essential
	EGF (human, recombinant)		5 ng/ml	optimal/beneficial
	bFGF (human, recombinant)		5 ng/ml	optimal/beneficial
Neuronal Cells				
FreeAdd 1X, 10%	EGF (human, recombinant)		50 ng/ml	optimal/beneficial
DMEM high glucose (ref.BW: SLD-527-500, SLD-525-500)		Insulin (recombinant human)	0.5 µg/ml	essential

* For correct cell attachment and spreading, addition of CaCl₂ (0.06mM) is also highly recommended.